

Strategic Salmon Health Initiative:

Defining the role of infectious disease in survival of migratory salmon and interactions with cultured salmon

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Oceans Canada



Challenges with our understanding of disease impacts on wild populations

- We rarely observe wild fish die, they simply disappear
- Acute versus chronic infections versus carrier states
- Many infectious agents never assessed in wild populations or in BC salmon, especially in the marine environment
- Potential for interactions between cultured and wild fish
- Impacts may vary by environment, species, and stocks
- Cumulative impact of stress and disease

Miller et al. 2014 Evolutionary Applications



Sub-lethal effects of infection may be more detrimental in wild than cultured fish



Swim performance
and predation



Feeding



Ion homeostasis



Behavioral shifts in
migration timing
and speed



Strategic Salmon Health Initiative

Discover the pathogens and potential diseases that may undermine the productivity and performance of BC salmon, their evolutionary history, and the potential role of exchanges between wild and cultured salmon

- Quantitative assessment of 47 potential pathogens in in >26,000 salmon
- Wild, enhancement hatchery, and farmed salmon
- Combines traditional and novel approaches to study disease
- Novel genomic technologies and systems biology approaches
- Considers both lethal and sub-lethal impacts of infection



Strategic Salmon Health Initiative

Reverse traditional approach starting with disease—

- 1) Broad-based quantitative infectious agent monitoring
- 2) Pathogenic potential
 - a) physiological impacts at molecular, protein and cellular levels
 - b) organismal impact via tracking, predation and holding/challenge studies
- 3) Novel pathogen discovery
 - a) Construction of disease networks to identify potential diseases with unknown etiological agents



High Throughput infectious agent monitoring

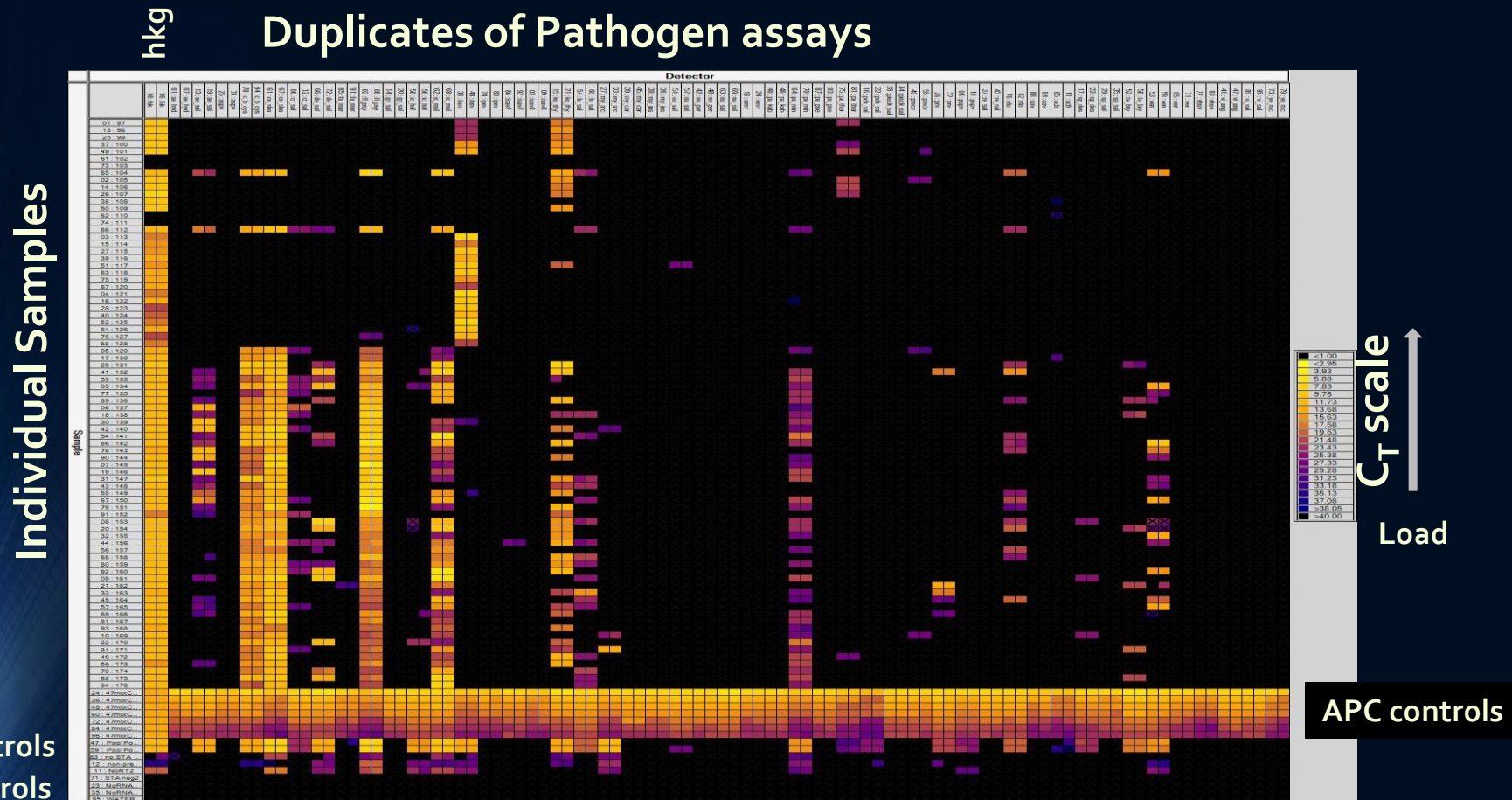


Identify the infectious agents detected in BC salmon, their temporal and spatial distributions, and the locations in which they are transmitted



Infectious agent monitoring heatmap

Duplicates of Pathogen assays



Salmon Pathogen Monitoring Platform

12 viruses

Virus	Abbreviation
Atlantic salmon paramyxovirus	aspv
Infectious hematopoietic necrosis virus	ihnv
Infectious pancreatic necrosis virus	ipnv
Infectious salmon anemia virus	isav7 / isav8
Oncorhynchus masou herpes virus	omv
Pacific salmon parvovirus	pspv
Piscine myocarditis virus	pmcv
Piscine reovirus	prv
Salmon alphavirus	sav
Viral erythrocytic necrosis virus	ven
Viral encephalopathy and retinopathy virus	ver
Viral hemorrhagic septicemia virus	vhsv

13 bacteria

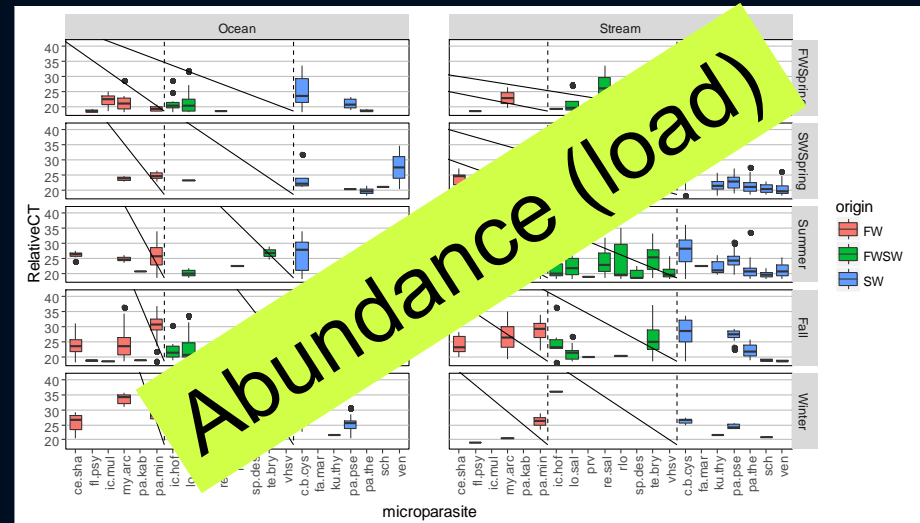
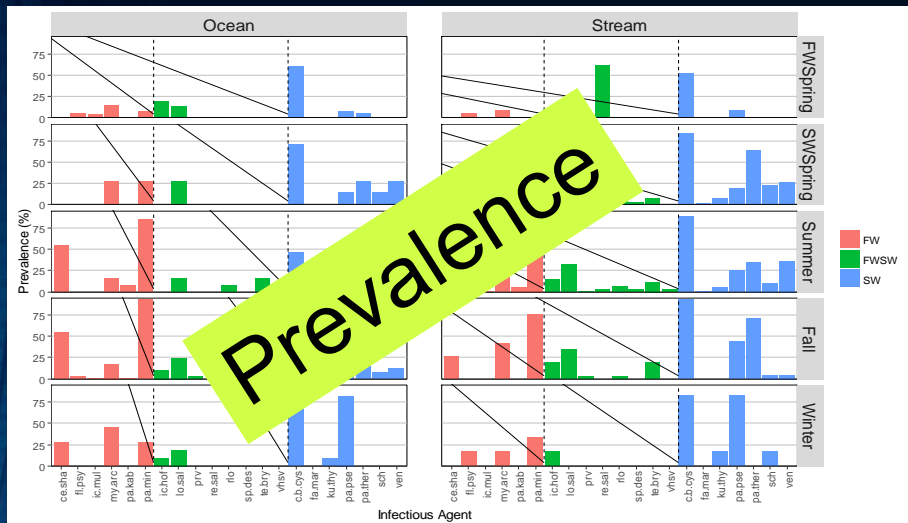
Bacterium	Abbreviation
<i>Aeromonas hydrophila</i>	ae_hyd
<i>Aeromonas salmonicida</i>	ae_sal
<i>Candidatus Branchiomonas cysticola</i>	c_b_cys
<i>Flavobacterium psychrophilum</i>	fl_psy
<i>Gill chlamydia</i>	sch
<i>Piscichlamydia salmonis</i>	pch_sal
<i>Piscirickettsia salmonis</i>	pisk_sal
<i>Renibacterium salmoninarum</i>	re_sal
Rickettsia-like organism	rlo
<i>Vibrio anguillarum</i>	vi_ang
<i>Vibrio salmonicida</i>	vi_sal
<i>Yersinia ruckeri</i>	ye_ruc
<i>Tenacibaculum meritimum</i>	Te_mer

- TaqMan assays to 47 agents suspected or known to cause disease in salmon world-wide
- 10 OIE-listed salmon pathogens

22 parasites

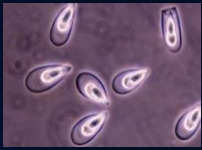
Parasite	Abbreviation
<i>Ceratomyxa shasta</i>	ce_sha
<i>Cryptobia salmositica</i>	cr_sal
<i>Dermocystidium salmonis</i>	de_sal
<i>Facilispora margolisi</i>	fa_mar
<i>Gyrodactylus salaris</i>	gy_sal
<i>Ichthyophonus hoferi</i>	ic_hof
<i>Ichthyophthirius multifiliis</i>	ic_mul
<i>Kudoa thyrsites</i>	ku_thy
<i>Loma salmonae</i>	lo_sal
<i>Myxobolus arcticus</i>	my_arc
<i>Myxobolus cerebralis</i>	my_cer
<i>Myxobolus insidiosus</i>	my_ins
<i>Nanophyetus salmincola</i>	na_sal
<i>Neoparamoeba perurans</i>	ne_per
<i>Nucleospora salmonis</i>	nu_sal
<i>Paranucleospora theridion</i>	pa_ther
<i>Parvicapsula kabatai</i>	pa_kab
<i>Parvicapsula minibicornis</i>	pa_min
<i>Parvicapsula pseudobranchicola</i>	pa_pse
<i>Sphaerothecum destructuens</i>	sp_des
<i>Spironucleus salmonicida</i>	sp_sal
<i>Tetracapsuloides bryosalmonae</i>	te_bry

Infectious agent monitoring identifies the shifting prevalence, abundance (load), and diversity of infectious agents through smolt and adult migration



FW Chinook Smolt Infectious Agent Detections

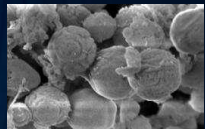
SW



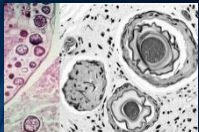
Parvicapsula



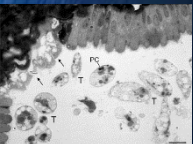
Ichthyophthirius



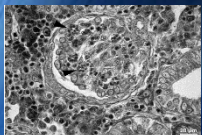
Sphaerothecum



Ichthyophonus



Ceratomyxa



Parvicapsula

24 of 45 agents with prevalence >1%

8 agents prevalence of <1%

16 of 32 agents originate in freshwater (FW)

5 viruses
8 bacteria
19 parasites

52 fish "agent-free" (most FW)

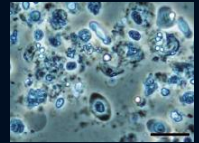
Infectious agent richness increases from FW
(average 1.5 agents/fish) to SW (average 4
agents/fish)

Maximum agent richness was 10/fish

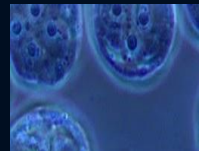
Strahan Tucker



Paranucleospora



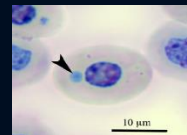
Parvicapsula



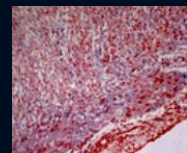
Tetracapsuloides



Rickettsia



VEN



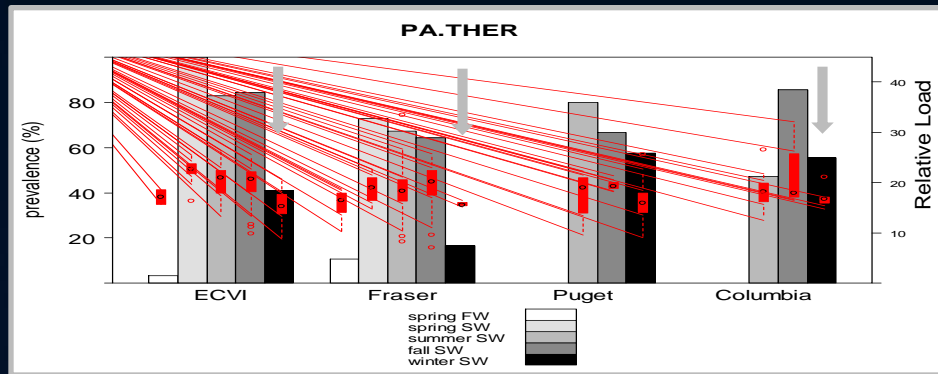
PRV

Agents Commonly observed in Salish Sea Chinook Salmon

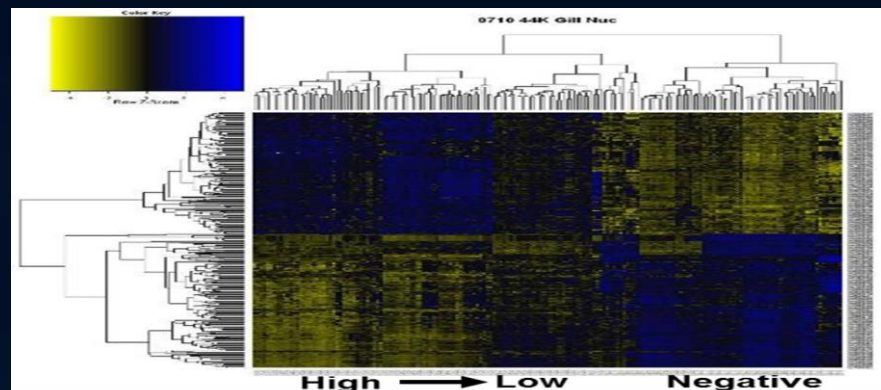
Paranucleospora theridion

SW microsporidian transmitted through sea lice

Proliferative gill disease (PGD) in European salmon



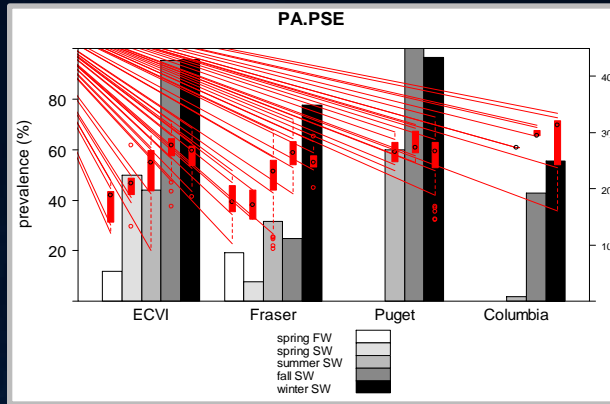
Sharp reduction in prevalence/load in winter consistent with timing of disease Oct-Feb in Europe



Powerful stimulation of immune response in Pacific salmon smolts

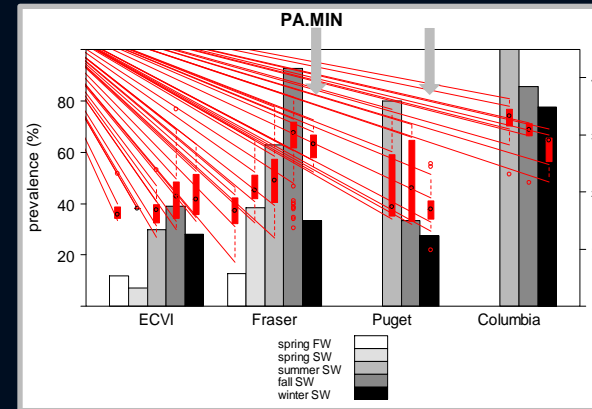
Microbes Commonly observed in Salish Sea Chinook Salmon

Parvicapsula pseudobranchicola
SW-transmitted myxozoan
PGD, vision impairment and anorexia in
European salmon

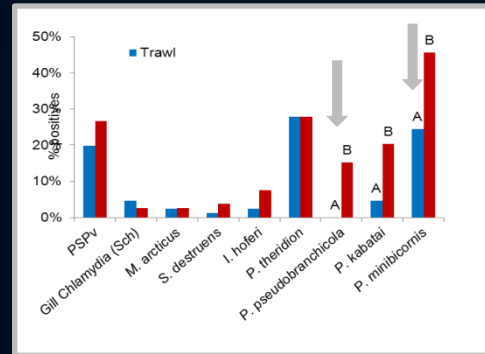


High prevalence and load fall/winter

Parvicapsula minibicornis
FW- transmitted myxozoan
Kidney disease



High prevalence/load Fraser- Columbia
Reduced prevalence and load truncation winter



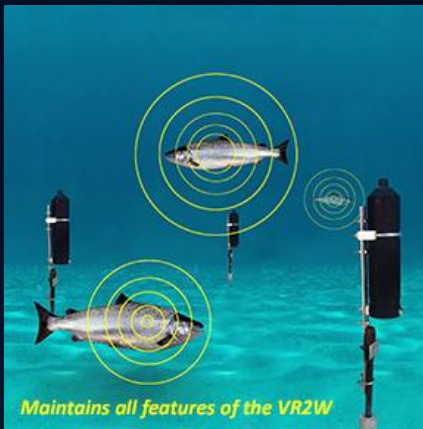
Increased Auklet predation risk in QC Strait sockeye

Linking infectious agents with disease



Which infectious agents carried by migratory salmon are actually impacting them?

Establishing Linkages with Survival



Tracking Studies



Predation Studies

But... linking pathogens with disease in migratory fish is more difficult

Acoustic Tracking Studies

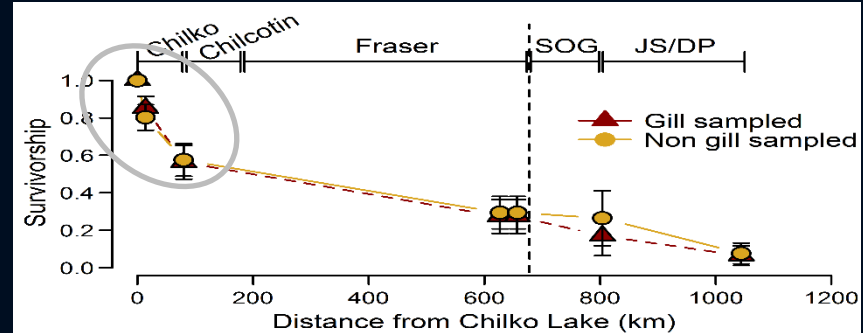
Downstream Migration Survival in Chilko Sockeye Smolts



Ken Jeffries

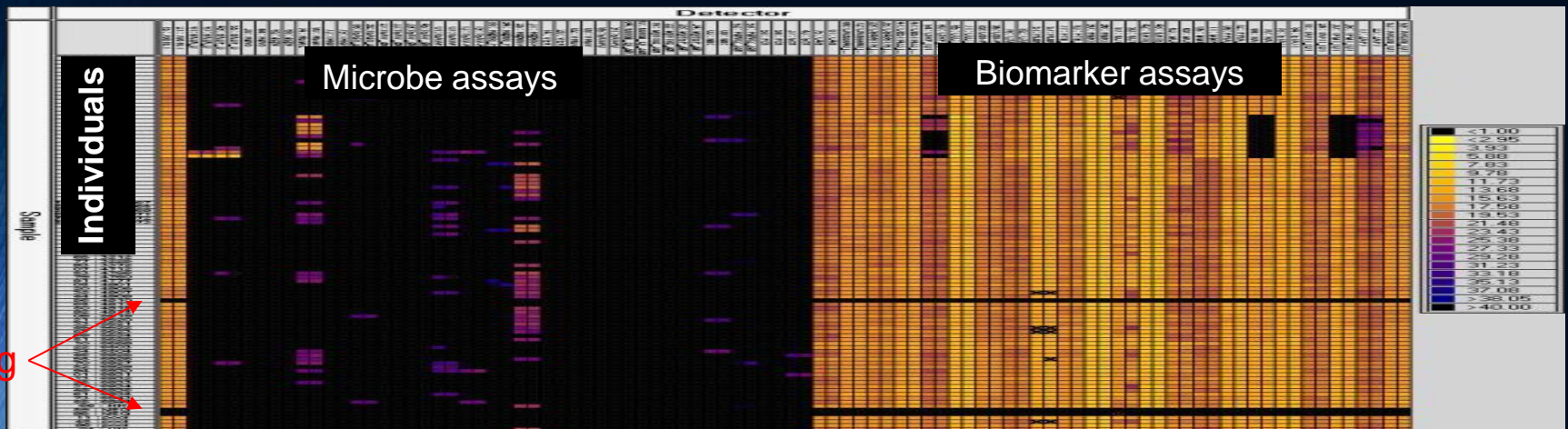


Chilko Smolt



40% "Immediate Mortality" in the Chilcotin

Merging microbe monitoring with host gene expression profiling



Powerful means to quickly identify microbes associated with immune stimulation, indicative of the potential for host damage/pathogenicity

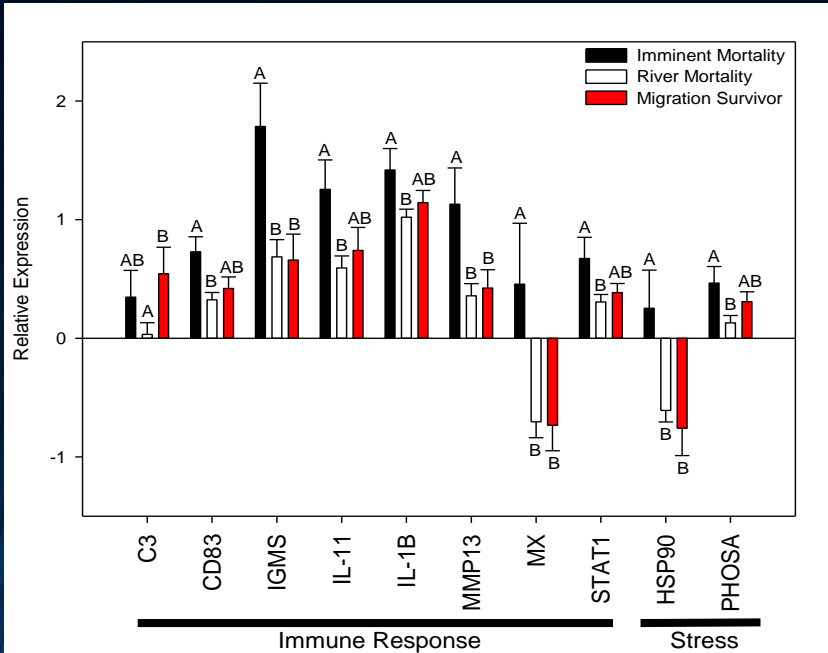
Jeffries et al. 2014
Molecular Ecology

Acoustic Tracking Studies

Downstream Migration Survival in Chilko Sockeye Smolts



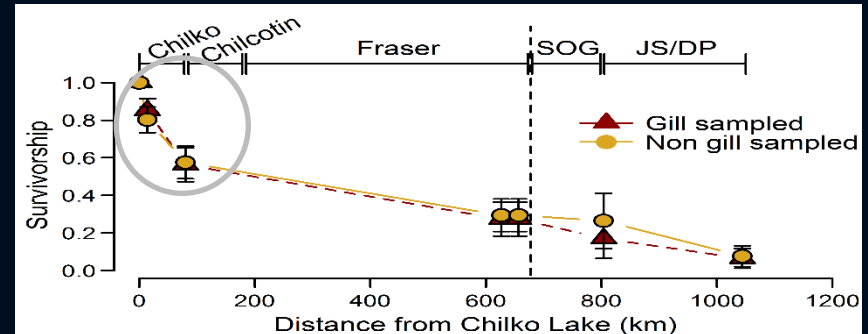
Ken Jeffries



10 (of 50) immune and stress genes associated with imminent mortality in the Chilcotin River

Anti-viral
T-cell activity
Antibody
Inflammation
Stress

Infectious Hematopoietic Necrosis Virus
Associated with stimulated immune-related signature and high, early losses of smolts



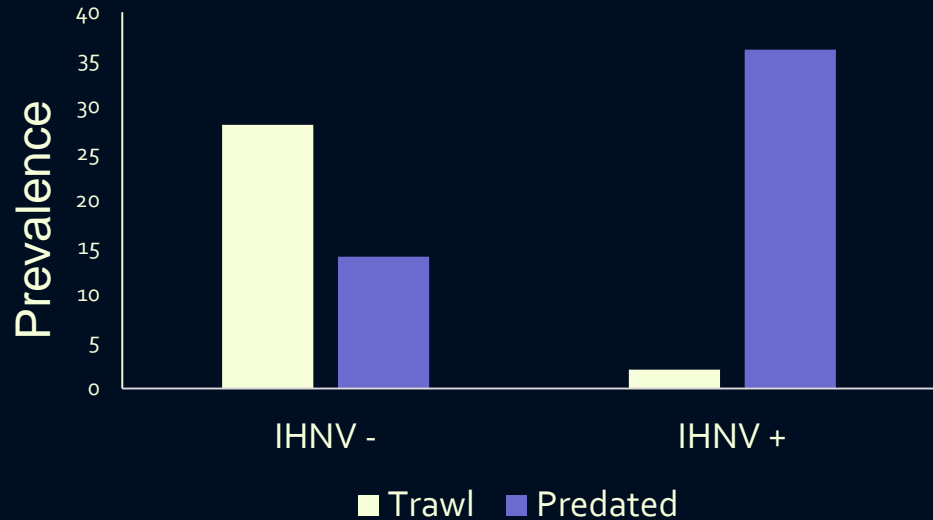
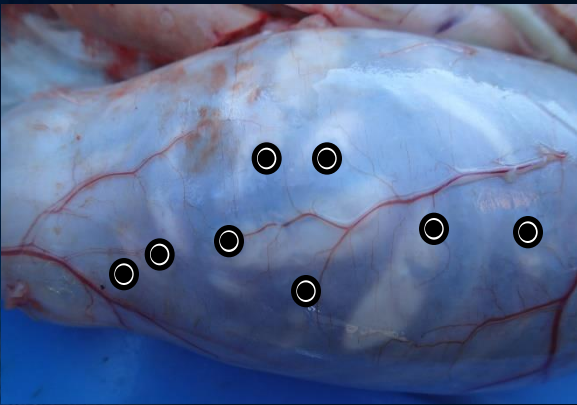
Fish with high IHNV loads disappear immediately and most with low-moderate loads don't make it to the ocean

But how?...

Jeffries et al. 2014
Molecular Ecology

Predation Studies

Bull trout predation during downstream migration in Chilko Sockeye smolts



Chilko smolts with IHNV had **34 times greater odds of being consumed** by bull trout than negative fish

Predation decreased the infective burden in the populations—thereby increasing population health

Nathan Furey
In prep



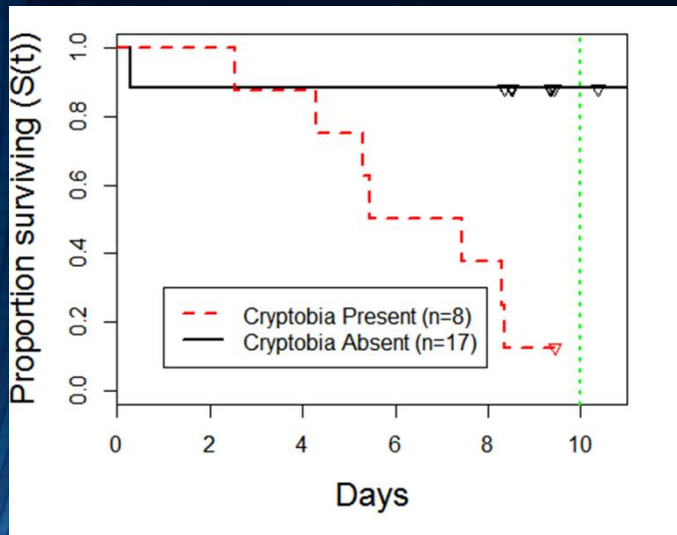
Holding Study

Adult salmon

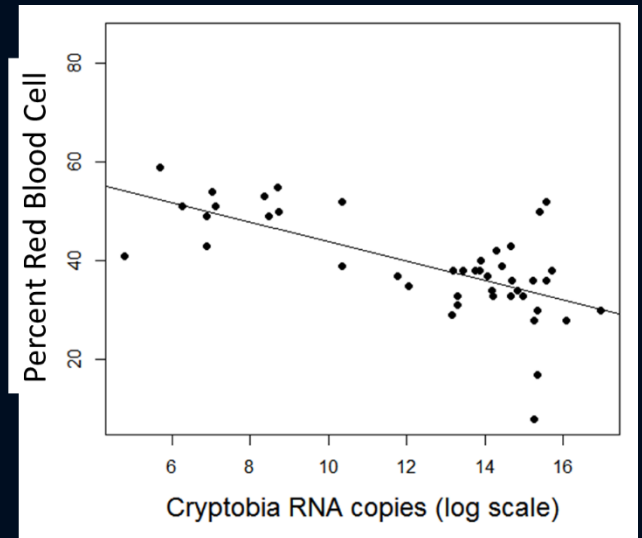
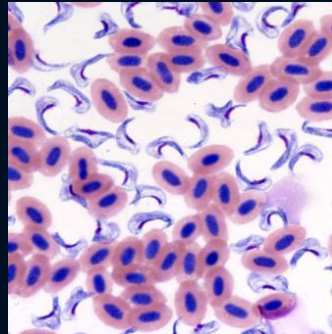
2013
Chilliwack River
Chinook Salmon



Amy Teffer



C. salmositica infection
reduced survival



C. salmositica infection impacts
blood physiology

Cryptobia salmonistica
FW Transmitted
Blood Flagellate Causing Anemia

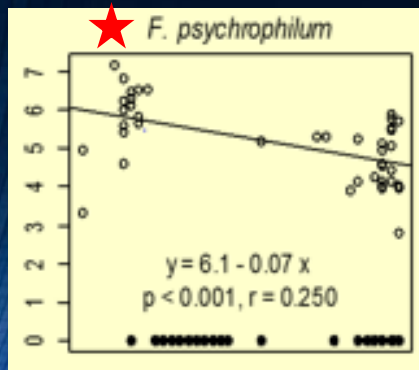
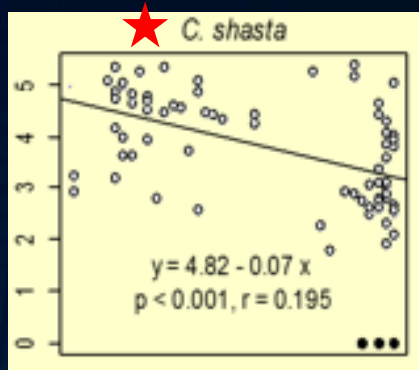
Amy Teffer
In prep

Microbes impacting fate associated with negative physiological impacts at molecular level



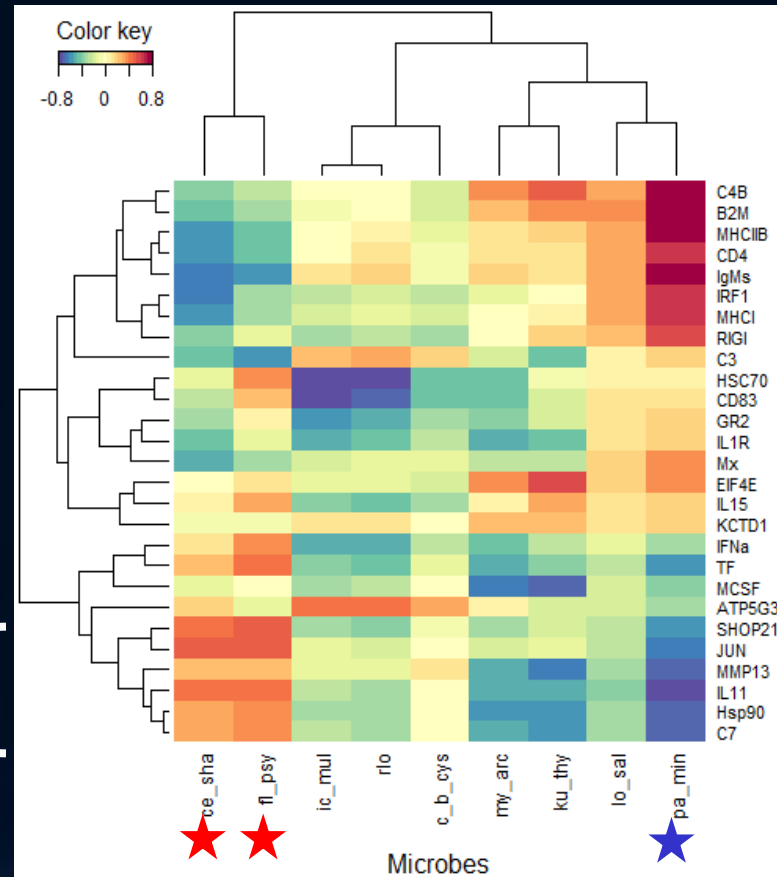
Amy Teffer

Early Stuart Adult Sockeye Salmon Holding Study



Days survived

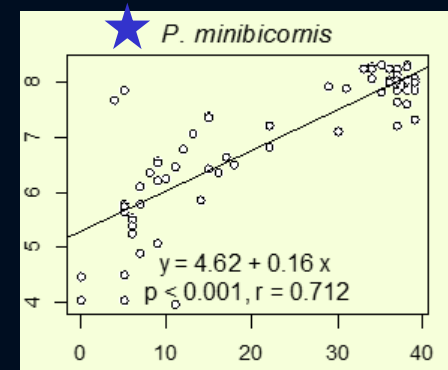
Early Mortality:
Stress, inflammation,
tissue damage,
immunosuppression



Transcriptional Signatures

Yersinia salmonae

Delayed Mortality:
Adaptive Immunity
Antibody production



Amy Teffer
In prep

Aquaculture Collections

Longitudinal Farm Sampling:

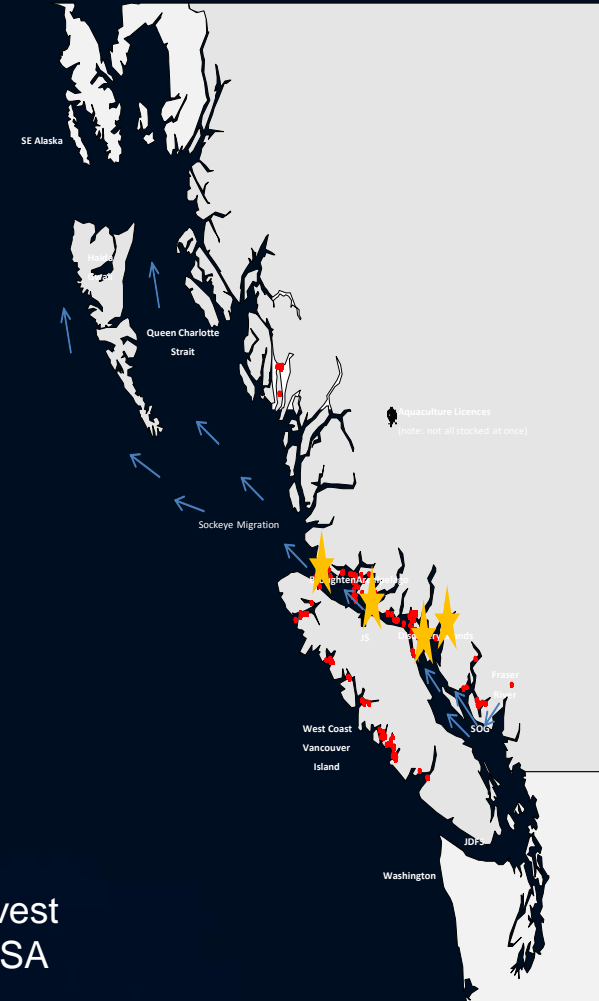
Healthy and moribund aquaculture fish from 4 ocean farms sampled regularly throughout ocean production cycle (~2500 fish)—healthy comparators with wild fish

Audit program

930 samples from 2011-2013 provide larger spatial/temporal scale to assess microbes associated with dying salmon



Working with Cermaq and Marine Harvest
Material Transfer Agreement with BCFSa



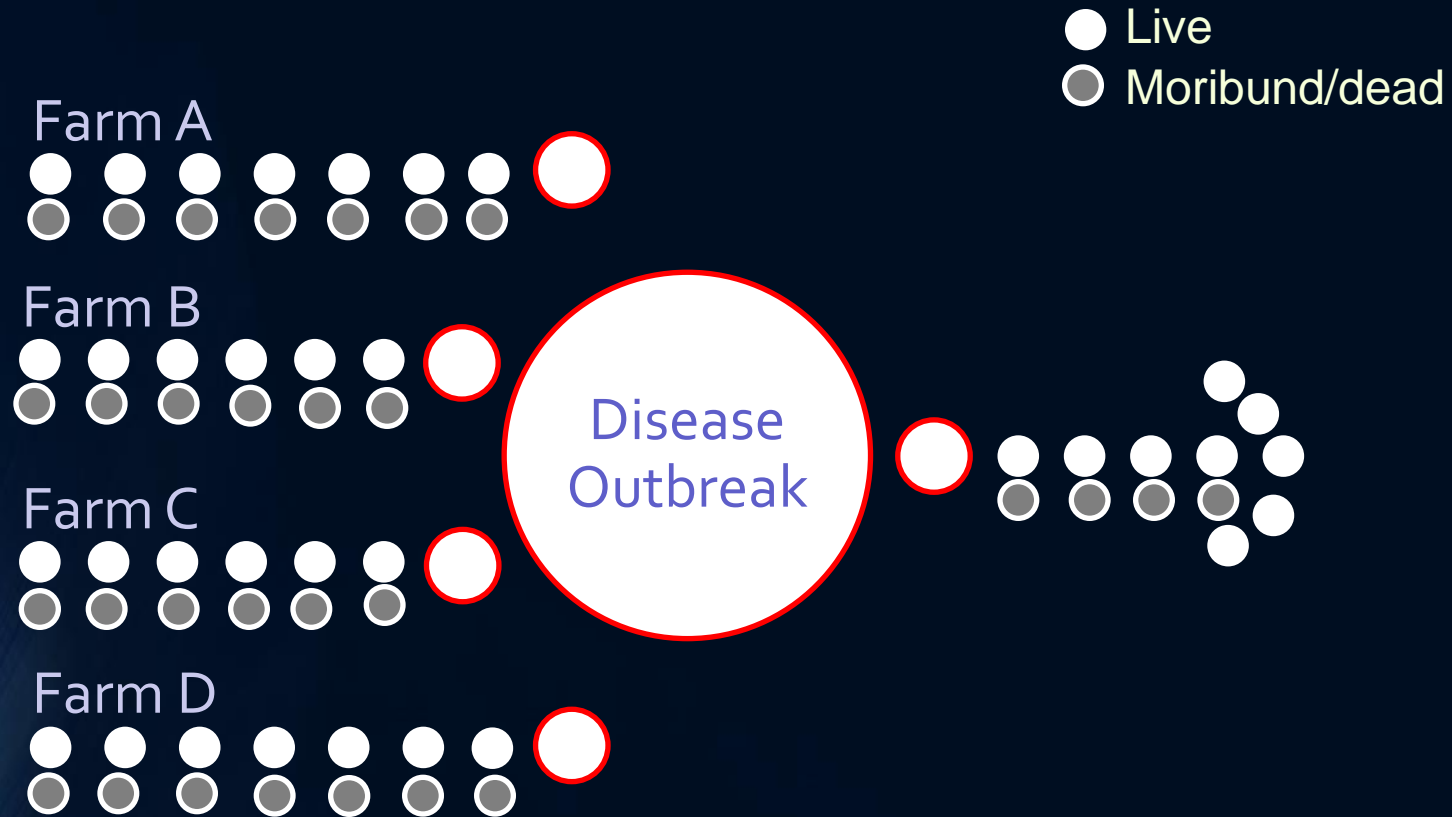
Longitudinal Farm Study Identifies Emerging Salmon Disease



Clinical pathology
Cellular pathology
Molecular profiling
Pathogen monitoring

Di Cicco et al. 2017
PLOS ONE

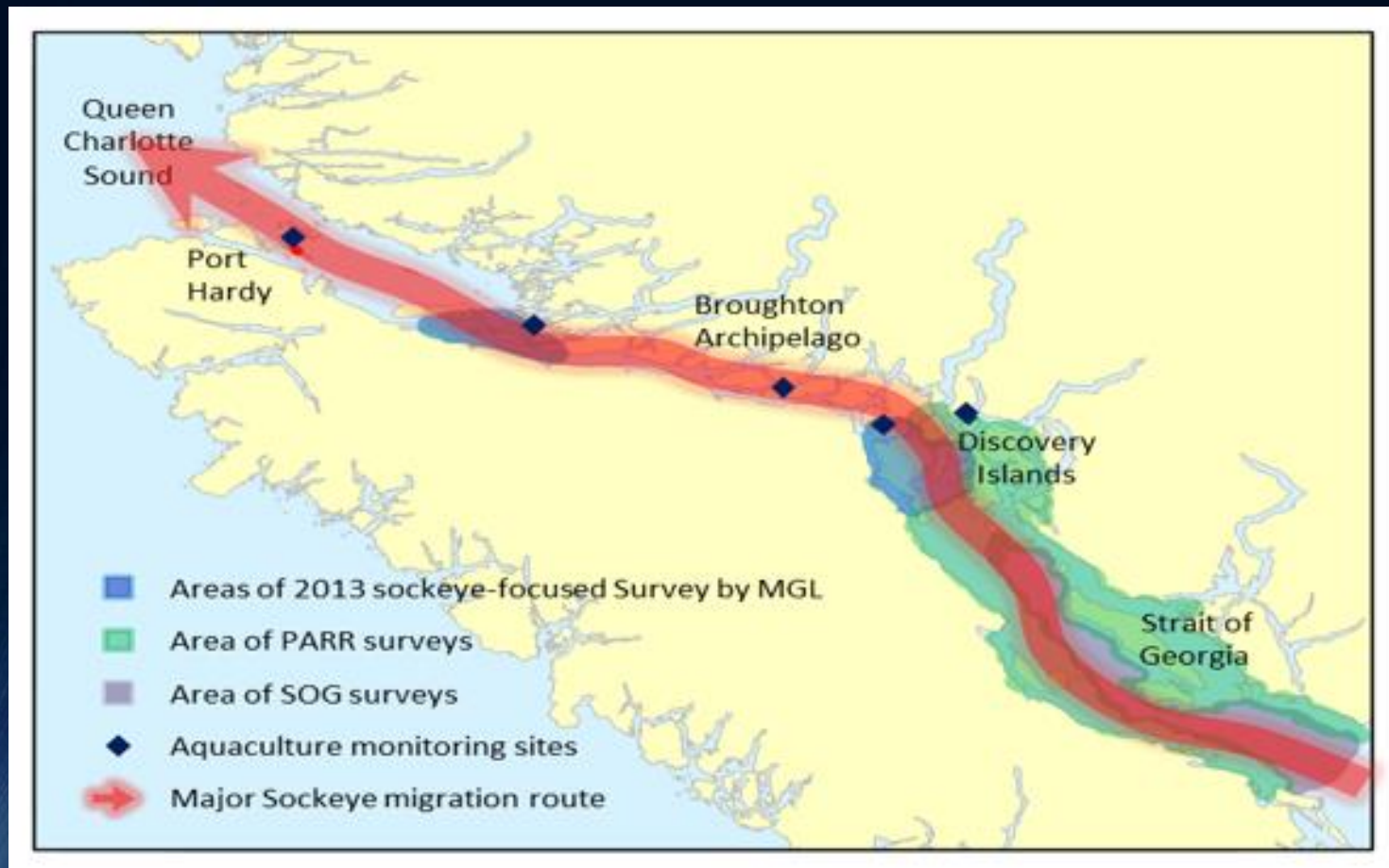
SSH Longitudinal Farm Sampling



Fine-scale temporal sampling to uncover cellular and molecular processes associated with disease development and recovery

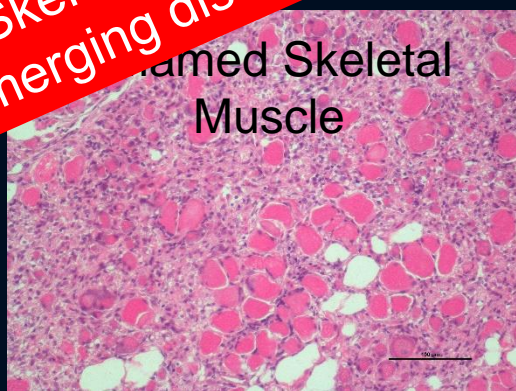
Collection of live fish provides best comparator to samples of migratory salmon

Aquaculture-Wild Interactions

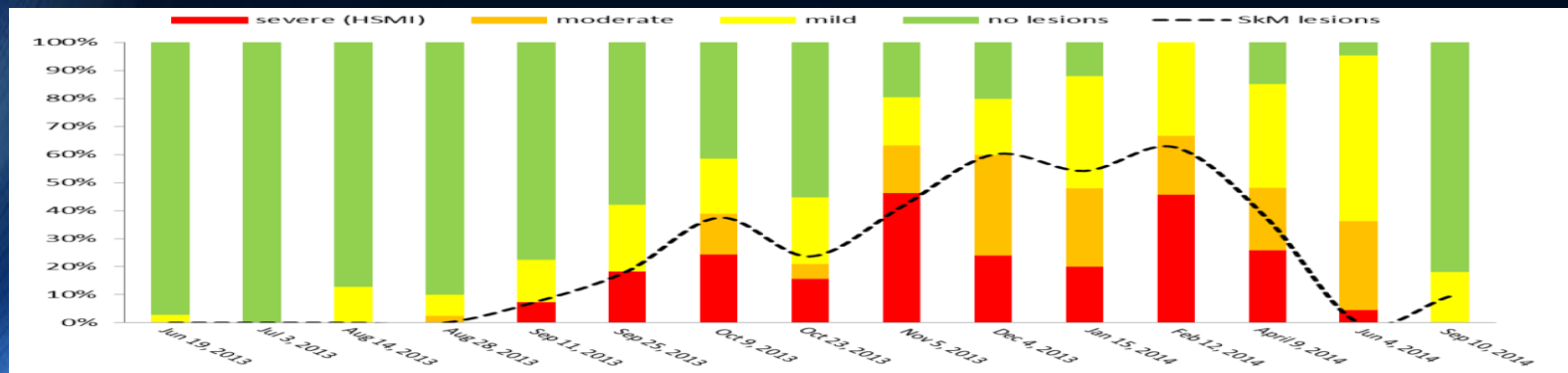


Sampled four geographically dispersed farms over the entire ocean production cycle along the migration pathway of wild salmon emanating from the Fraser River

Histopathological Investigations – Farm A



Diagnostic of Heart and Skeletal Muscle Inflammation (HSMI) disease
Third most impactful emerging disease in farmed Norwegian salmon

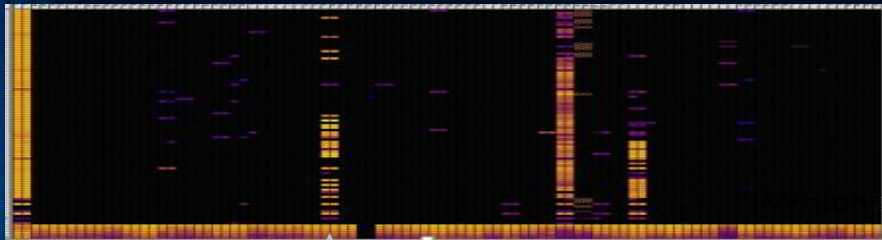
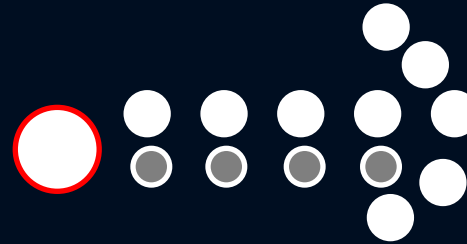


Longitudinal study resolves full developmental pathway of HSMI

Farm A

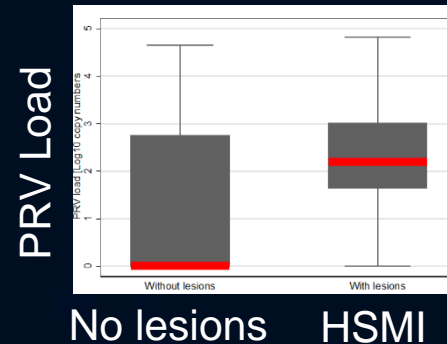


Disease
Outbreak

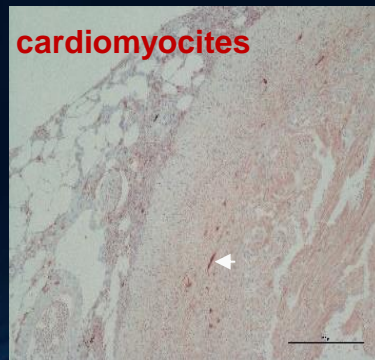
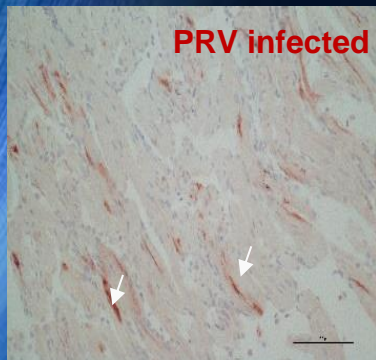


★ PRV P. ther Kudoa

High throughput pathogen monitoring:
Identify the shifting pathogen distributions in the heart



Epidemiological
Analyses:
Identify **PRV Pathogen loads** correlated with **lesion scores**



Immunohistochemistry:
Localization of PRV pathogen within the area of tissue damage

Longitudinal study resolves full developmental pathway of HSMI

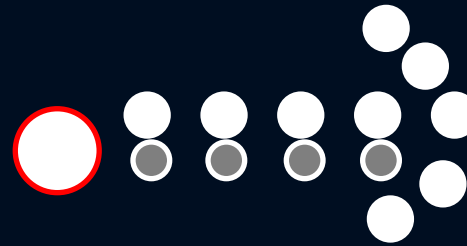
Farm A



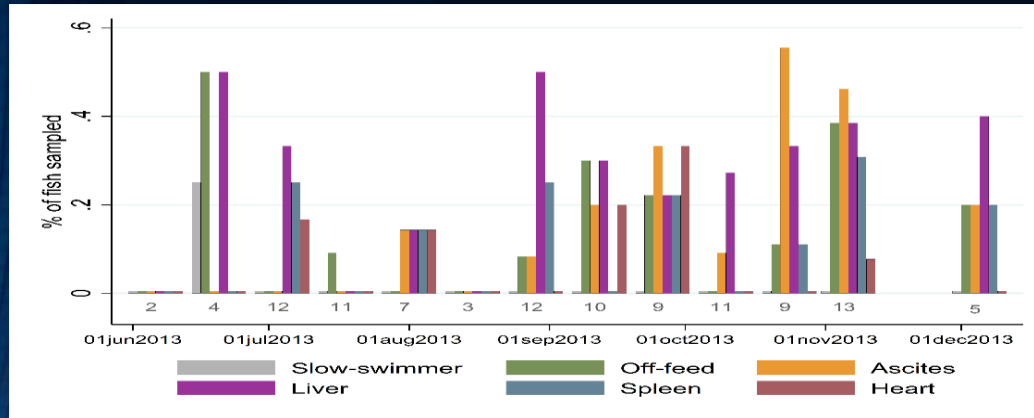
Mild lesions

Strong lesions

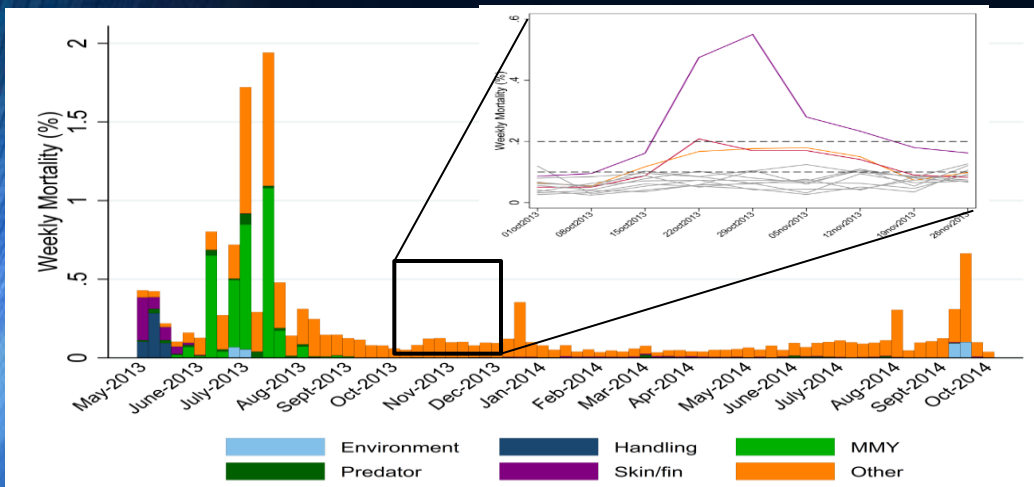
Disease
Outbreak



Live
Moribund



Clinical data:
Consistent with HSMI outbreaks in Norway



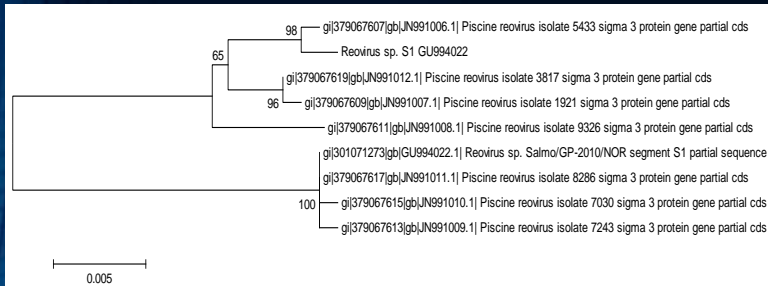
Mortality data:
Minimal impact on survival of fish on the farm

Longitudinal study resolves full developmental pathway of HSMI

Farm A



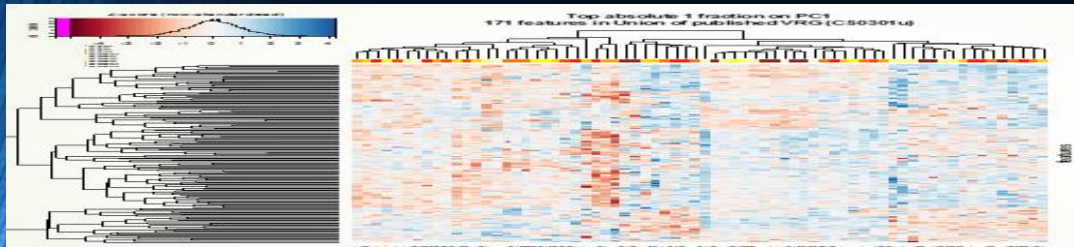
Disease
Outbreak



High throughput sequencing:

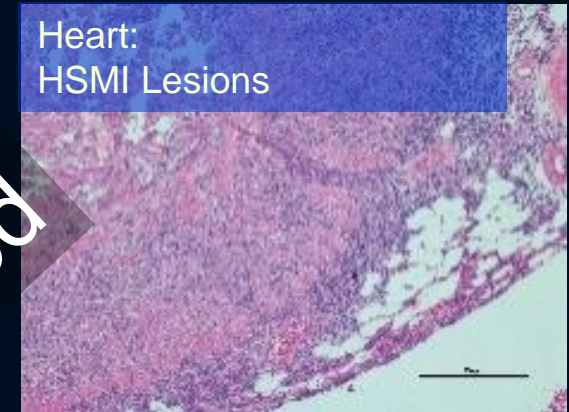
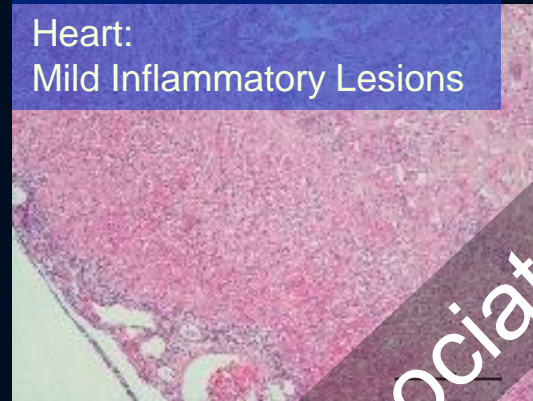
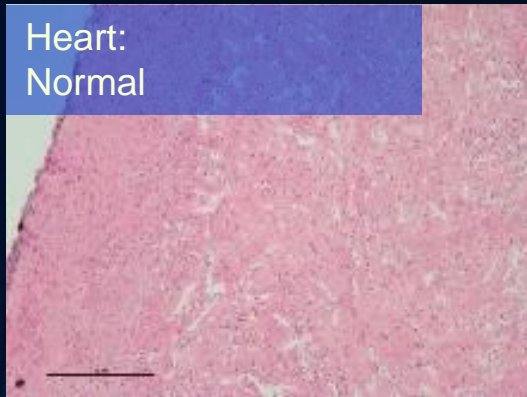
Full viral genome sequencing identifies PRV strain 99.9% similar to sequences previously observed in wild-migrating BC salmon

Identifying viral transcriptome shifts over disease cycle

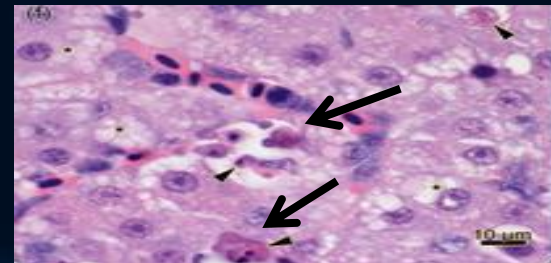
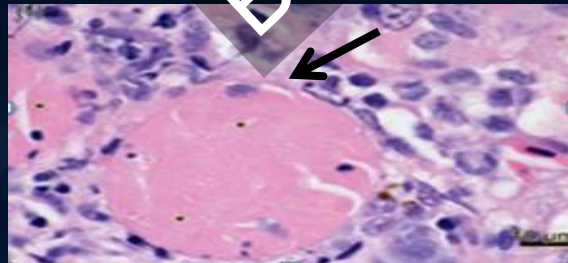
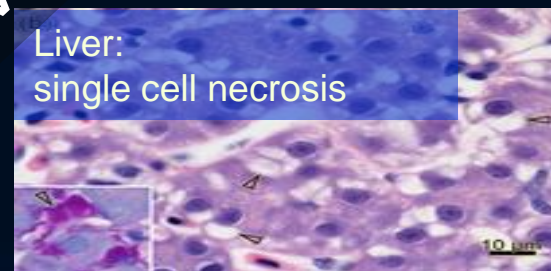
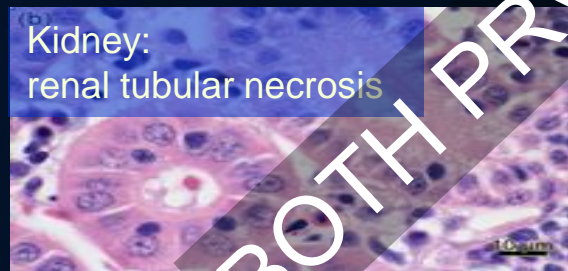


Transcriptomics (RNA-seq) (Underway):
Does the transcriptional profile match HSMI in Norway?

Viral Diseases in BC Farmed Salmon



Heart and Skeletal Muscle Inflammation in Atlantic Salmon



Jaundice Syndrome in Chinook Salmon

PRV highly prevalent in farmed fish (~70% of farm audit samples)

Virus increases in prevalence over the first 6 months in the ocean, while the two diseases associated with it occur ~8 months post ocean-entry



PRV detected, but NOT common, in migratory smolts

Virus increases from summer through fall/winter in Chinook and Sockeye salmon; 7% overall in Chinook and 3% in Sockeye



Our program has NOT detected viruses

ISAv – Infectious Salmon Anemia virus

IPNV – Infectious Pancreatic Necrosis virus

Omv -- Oncorhynchus Masu Herpesvirus

Sav – Salmon Alphavirus

PMCV – Piscine Myocarditis Virus

ASPV – Atlantic Salmon Paramyxovirus



Infective agents can be present in the absence of disease

- Farmed salmon
- 39% of Atlantic salmon and 52% of Chinook salmon with high loads of *Renibacterium salmoninarum* diagnosed with BKD
- 58% of Atlantic salmon and 31% of Chinook salmon with high loads of *Piscirickettsia salmonis* diagnosed with Rickettsiosis
- 35% of Atlantic salmon with high loads of *Tenacibaculum maritimus* diagnosed with Mouth Rot
- 30% of Chinook salmon audits with high PRV loads diagnosed with Jaundice/anemia
- 13% of Atlantic salmon audits with high PRV loads diagnosed with HSMI



How do we link PRV and other potential pathogens with disease if migratory fish disappear soon after they are physiologically compromised?



THE ANSWER:
Develop molecular methods to recognize early developing disease states

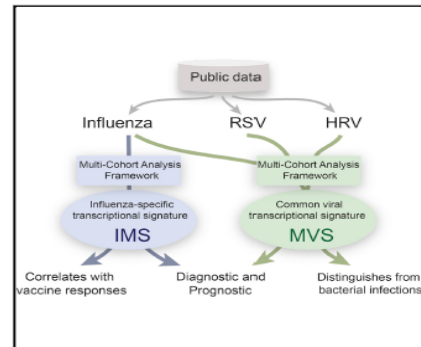


Molecular Disease Diagnostics – human medicine

Immunity

Integrated, Multi-cohort Analysis Identifies Conserved Transcriptional Signatures across Multiple Respiratory Viruses

Graphical Abstract



Authors

Marta Andres-Terre, Helen M. McGuire, Yannick Pouliot, Erika Bongen, Timothy E. Sweeney, Cristina M. Tato, Purvesh Khatri

Correspondence
pkhatri@stanford.edu

In Brief

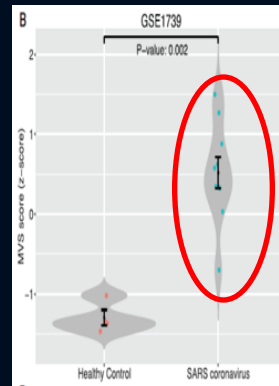
Clinically relevant respiratory viral signatures have not been defined. Khatri and colleagues identified host transcriptional responses common to multiple respiratory viruses (MVS) or specific to influenza (IMS) by leveraging heterogeneity present in public datasets. Both signatures distinguish viral from bacterial infections and IMS also distinguishes influenza from other viral infections.

Andres-Terre et al., 2015, *Immunity* 43, 1199–1211
December 15, 2015 ©2015 Elsevier Inc.
<http://dx.doi.org/10.1016/j.immuni.2015.11.003>

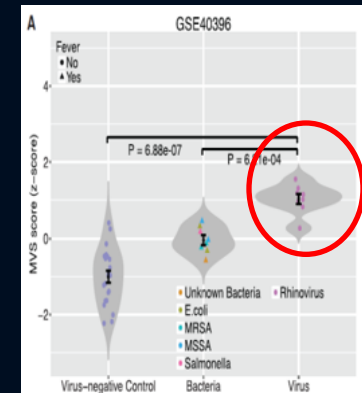
- Mined public transcriptome studies
- Identified diagnostic biomarker signatures for
 - respiratory viral disease
 - specific to influenza virus disease

Discovery

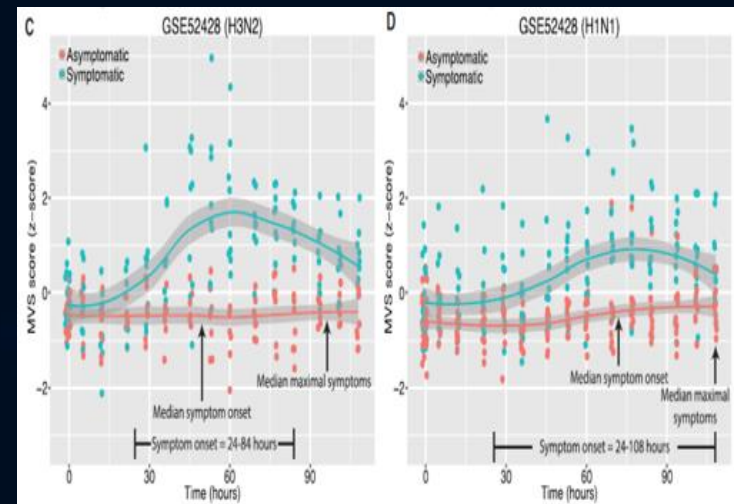
Validation



viral vs. healthy



viral vs. bacterial



Pre-symptomatic viral disease development

Viral Disease Development [VDD]

Salmon RNA viruses

VDD – predictive for all viruses

Multiple viruses

Krasnov
2011

Skjseol
2011

LeBlanc
2010

ISAV PMCV PRV IPNV
Atlantic

IPNV
Atlantic

ISAV
Atlantic

Published signatures

Union (532 features)



GUNTHER ANALYTICS
Data Analysis, Modeling and Simulation

Multiple salmon species

Exploration Analysis

Gene Shaving, Sparse Independent PCA

Purcell
2011

Miller
2007

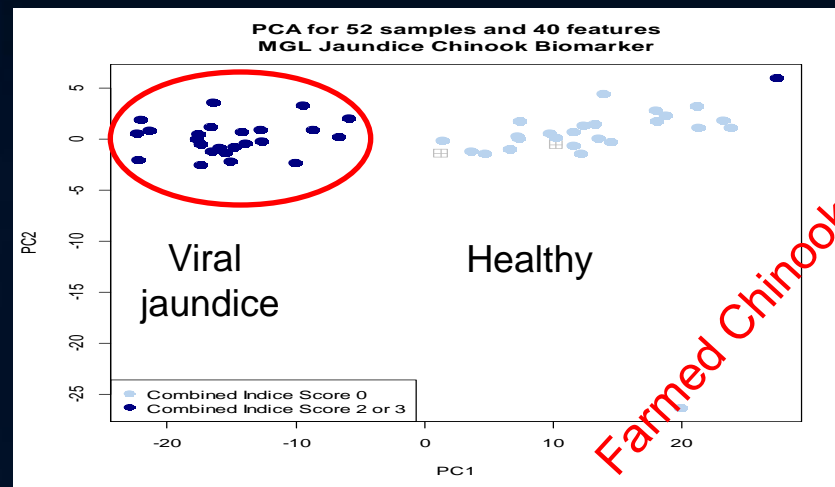
IHN
Rainbow Trout

IHN
Sockeye Chum
Atlantic Chinook

VDD 44 biomarker panel

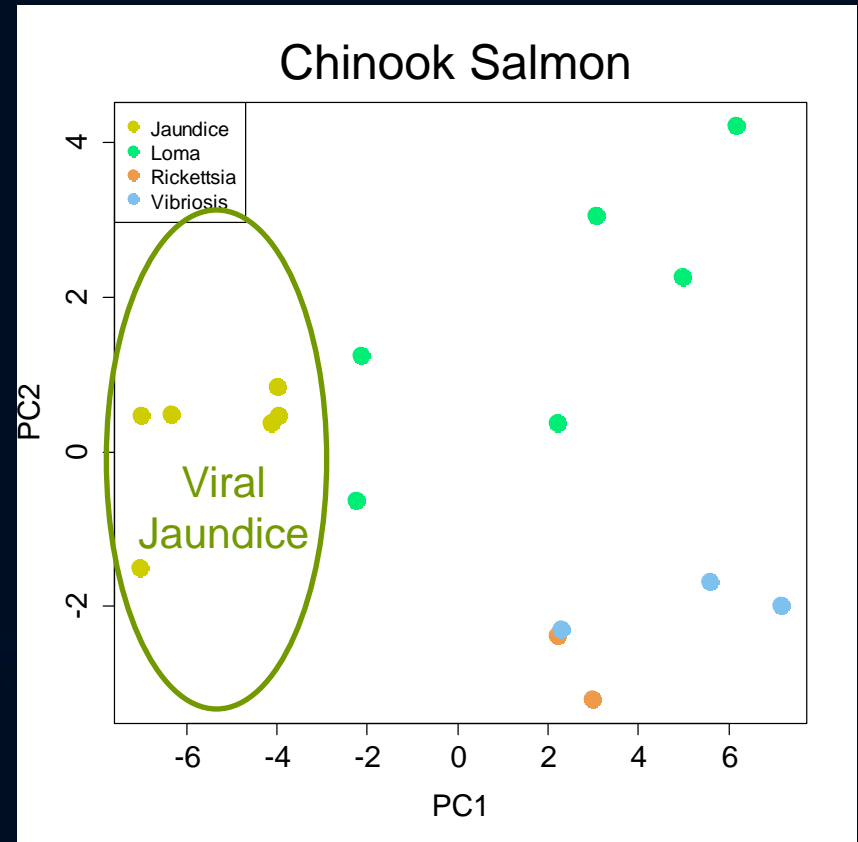
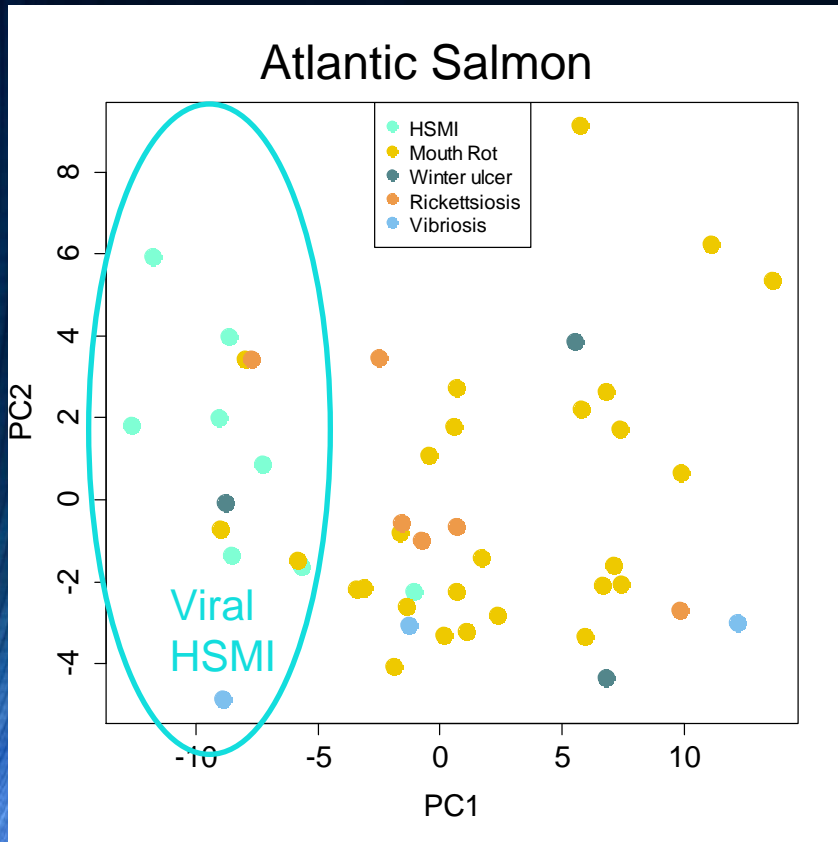
Molecular biomarkers – early disease detection

Molecular biomarkers identify development of viral disease state



Molecular biomarkers – early disease detection

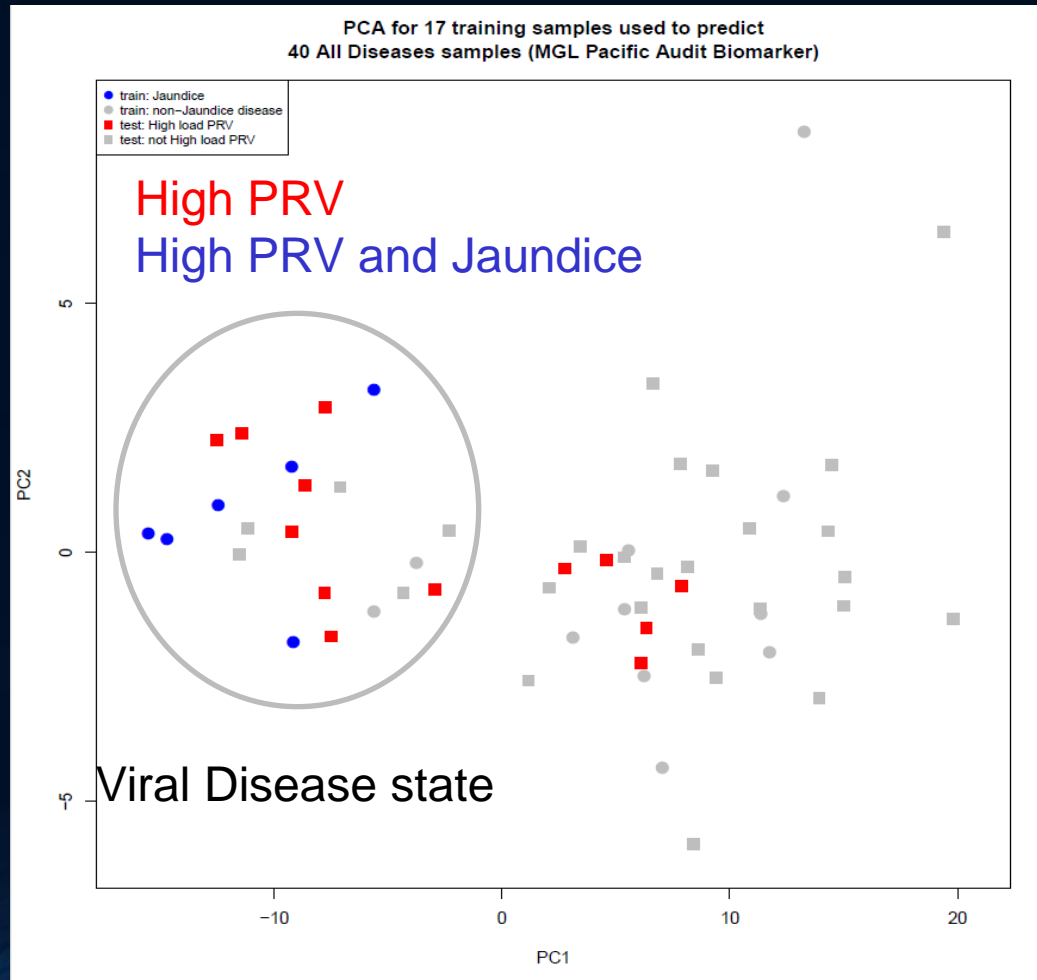
Molecular biomarkers identify differentiate fish with viral versus bacterial diseases



Mixed Tissues, Dead sampled fish,
Diagnosed through Veterinary Pathology

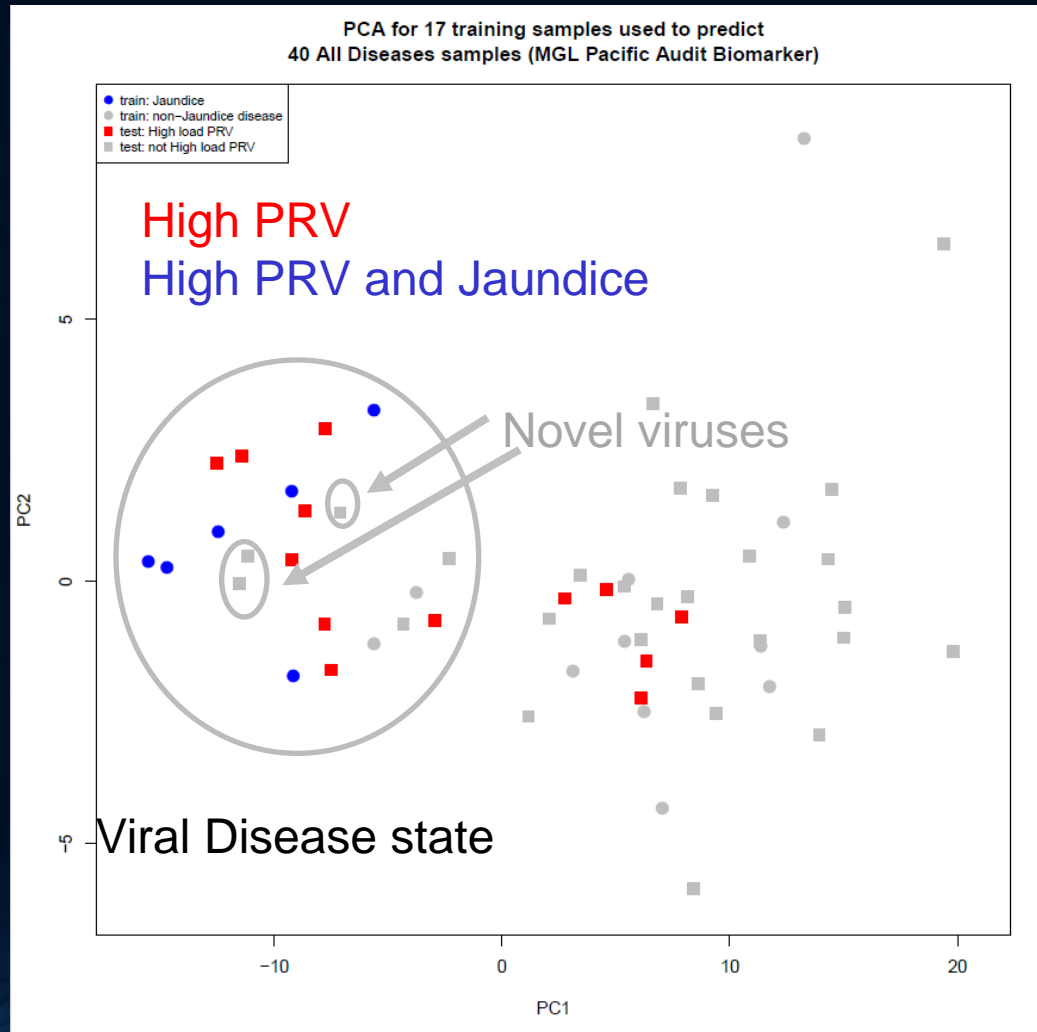
Molecular biomarkers – early disease detection

80% of Chinook salmon containing high loads of PRV are in a “viral disease state”
50% of which are diagnosed with jaundice/anemia
Wild Chinook salmon show the same 80% pattern

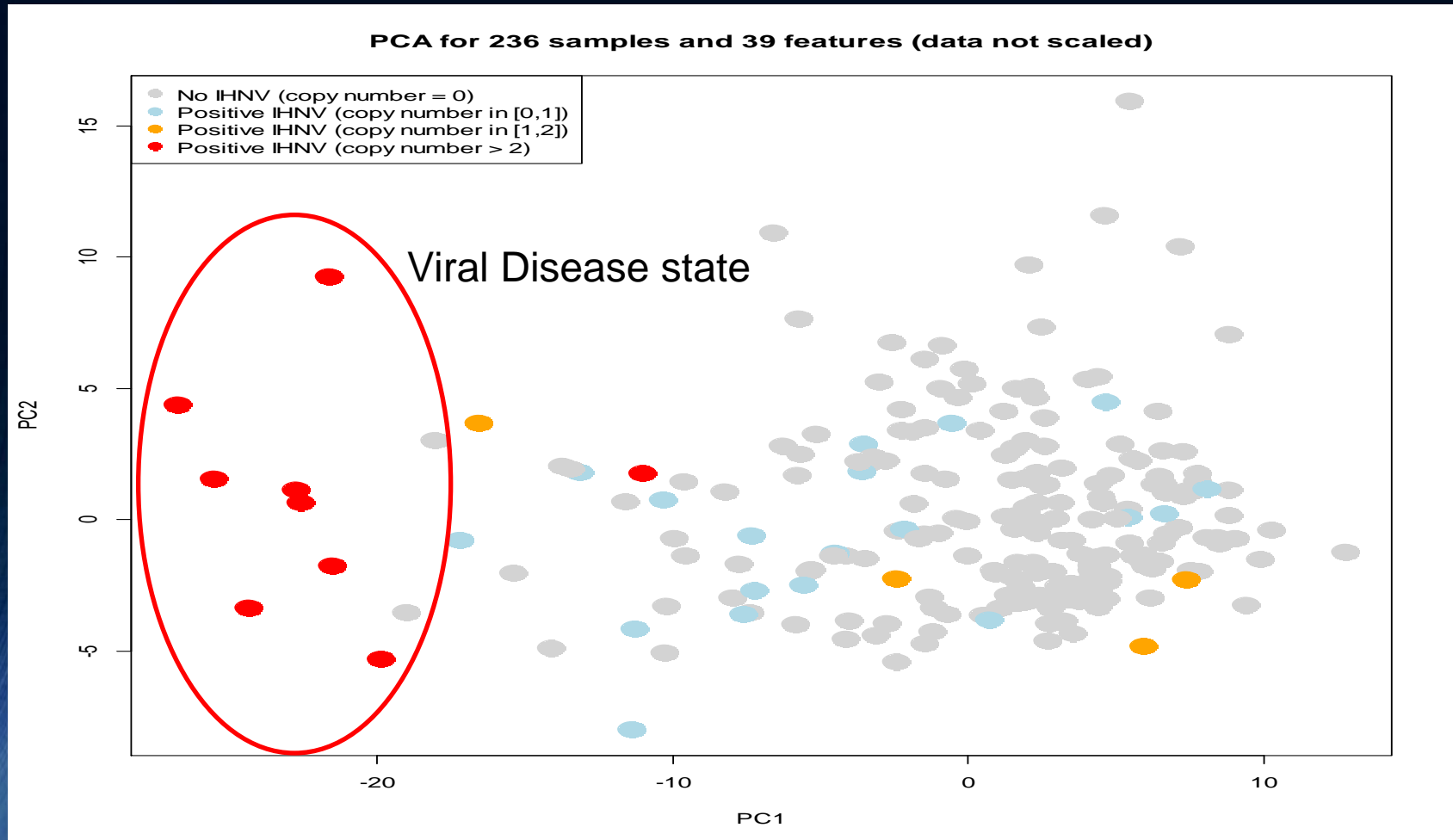


Molecular biomarkers – early disease detection

VDD Panel can be used to identify fish with uncharacterized viruses associated with a developing disease state



Wild Chilko Sockeye Smolts with high loads of IHNV in a viral disease state



- Move from “bug hunting” to “molecular disease diagnostics”
- Non-destructive gill biopsies — ideal for application with tracking studies

Key Findings to date

- Migratory salmon – natural exposure to wide array of potential pathogens
 - Half come from freshwater
 - An array of pathogens have already been associated with poor migratory success
 - Infection status enhances risk of predation
 - Fungal and protozoan parasites most commonly associate with poor survivorship outcomes
- Farmed salmon – The PRV-associated diseases HSMI and jaundice are present on BC salmon farms, but many other viral diseases are absent
 - We do not know:
 - Impacts on wild Pacific Salmon
 - How wide-spread
 - Industry-wide impacts on survival of farmed salmon
 - If factors in addition to PRV are required to initiate disease



SSHI: Linkages between Science and Regulation

- Develop innovative high throughput diagnostic tools for monitoring by regulators and industry
- Identify pathogens and diseases that pose biosecurity risks to Pacific salmon
- Contribute to DFO risk assessments associated with pathogen transmission from salmon farms and salmon enhancement hatcheries
- Inform disease management practices on farms and in salmon enhancement hatcheries
- Inform policies on minimal testing required to release smolts to the ocean
- Inform policies on the placement of farms within the migratory pathways of wild salmon





Strategic Salmon Health Initiative: Project Team Acknowledgements



Fisheries and
Oceans Canada

Genomics: Karia Kaukinen, Angela Schulze, Shaorong Li, Tobi Ming, Amy Tabata, Norma Ginther

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Epidemiology/Sequencing: **Raph Vanderstichel, Ian Gardner (UPEI), Curtis Suttle (UBC)**

Salmon Ecology / Physiology: **Scott Hinch, Tony Farrell (UBC), Stephen Cooke (Carlton), David Patterson, Strahan Tucker**

Students/PDFs: **Art Bass, Amy Teffer, Ken Jeffries, Nathan Furey, Aimee Lee Houde, Katrina Cook, Jacqueline Chapman, Christine Stevenson, Steve Healey, Yovela Wang, Gideon Mordecai, Emilie Lauren, Krishna Thacur, Dianna Jarmillo**

Project Leads: **Brian Riddell (PSF)** and Kristi Miller (DFO)

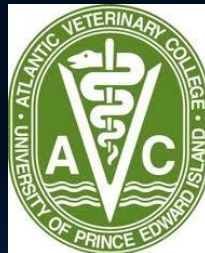


Questions?



Fisheries and Oceans
Canada

Pêches et Océans
Canada



CVER
UPEI Centre for Veterinary
Epidemiological Research

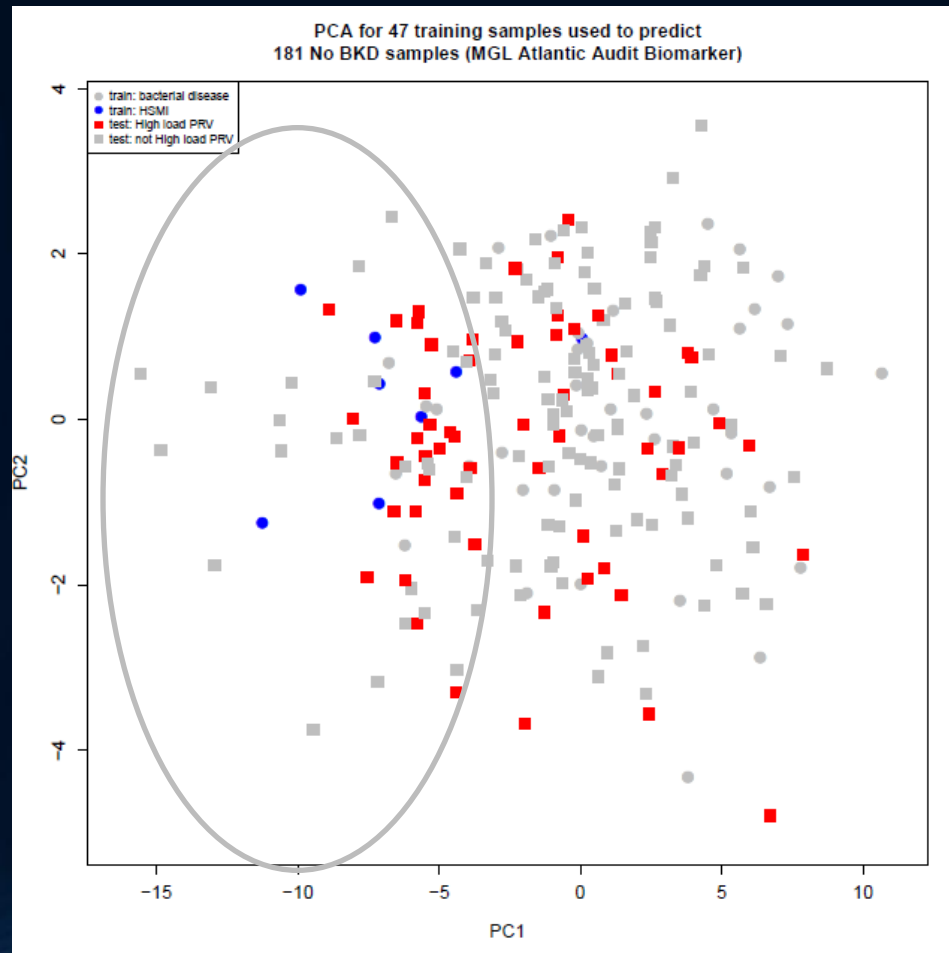


Molecular biomarkers – early disease detection

50% of Atlantic salmon containing high loads of PRV are in a “viral disease state”

25% of which are diagnosed with jaundice/anemia

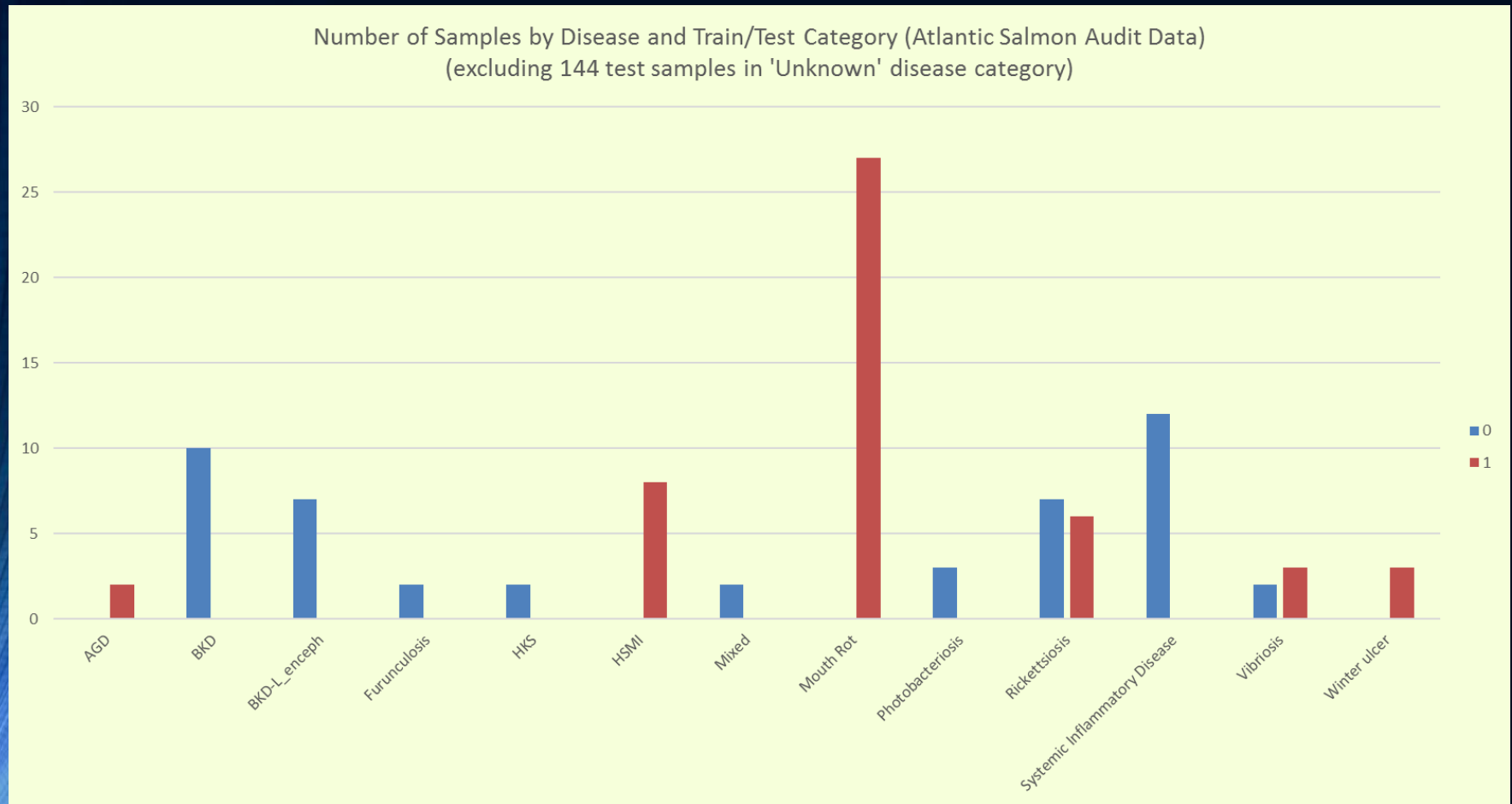
34% of dying chinook are in a VDD state—half with unknown viral associations



What do we know about PRV impacts on Pacific Salmon?

- Jaundice and anemia has been reported in farmed BC Chinook salmon, *Oncorhynchus tshawytscha*, coincident with high load PRV infections
- Jaundice and HSMI lesions associated with PRV infection has also been observed in farmed Rainbow Trout (Norway; Olsen et al. 2015) and Coho salmon (Chile – Godoy et al. 2016; and Japan – Takano et al. 2016).
- A BC challenge study failed to reproduce mortality, clinical signs, or the lesions found on fish dying of jaundice on the farms (Garver et al. 2015).
- However, the study identified mild lymphohistiocytic endocarditis lesions (early stage inflammation of the heart) exclusive to challenged fish and in all the three species tested (Chinook [60%], Sockeye [70%] and Atlantic [20%] salmon).
- Japanese study found cause and effect relationship between a novel strain of PRV and EIBS associated jaundice in Coho salmon (Takano et al. 2016)
- At this point, it would be remiss to discount these global findings in our assessments of “risk” of PRV to Pacific salmon

Diagnostic Distribution of Atlantic salmon audits



Diagnostic Distribution of Chinook salmon audits

