

Chronic Wasting Disease (CWD) in British Columbia

Instructions below are presented as a guide to identifying, sampling, and submitting the correct tissues for chronic wasting disease (CWD) testing in cervids (deer, elk, caribou, and moose).

A Note on Safety

Use a sharp knife and disinfect all tools and surfaces before and after use with a 40% bleach solution. Although there have been no confirmed cases of CWD in humans, Health Canada recommends that animals positive for CWD should not be eaten.

CWD Submission Checklist

- Prepare sample in one of three ways:
 - Cut head off animal and remove antlers;
 - Cut out lower jaw with the tongue and tissues at back of throat;
 - Remove **lymph nodes** and **tonsils** (for deer), or **lymph nodes** and **obex** (for elk, moose, and caribou).
- Fill out CWD Ear Card provided at freezer.
- Retain perforated portion or take a photo of your Ear Card to look up your results online when they become available.
- Attach CWD Ear Card to sample with zip-tie (if submitting head or jaw) or in a ziplock bag (if submitting tissues).

Note: Do not attach Ear Card to outside of bag.

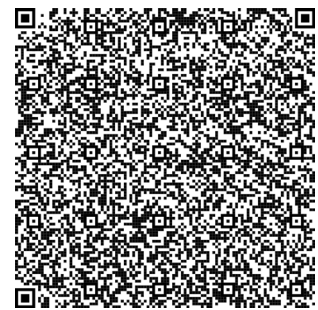
- Freeze sample and submit at one of our drop-off locations: gov.bc.ca/CWDdropoff
- Check results online using your unique CWD Ear Card number: gov.bc.ca/CWDresults

Ask a question about CWD:

Email: CWD@gov.bc.ca

Phone: 250-751-7246

For video instructions on how to sample a deer head for CWD:



Find out where to submit samples:

gov.bc.ca/CWDdropoff



Thank you for your participation.
Hunter-submitted samples are vital to CWD surveillance.

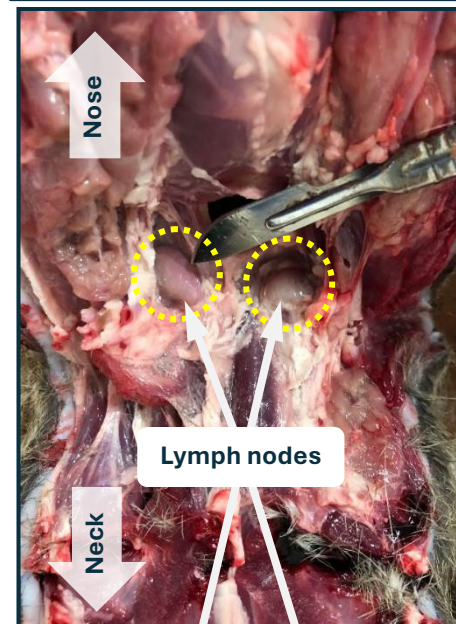
If you are sampling a **WHITE-TAILED DEER** or **MULE DEER**

Retropharyngeal Lymph Nodes (RPLNs)

1. Position head upside-down.
2. Using a sharp knife, cut underneath the jaw through the trachea and the muscles of the neck to the joint of the skull and the first vertebrae.
3. Lift the trachea, pull towards the nose of the animal, and cut away the muscles under the trachea until you reach fat pockets near the joint of the first vertebrae.
4. Cut through the fat pockets carefully to expose the left and right lymph nodes.

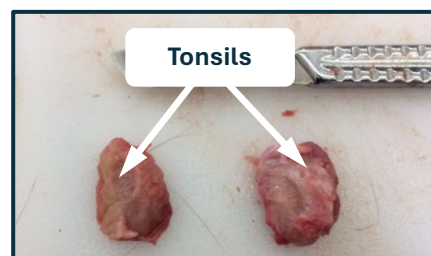
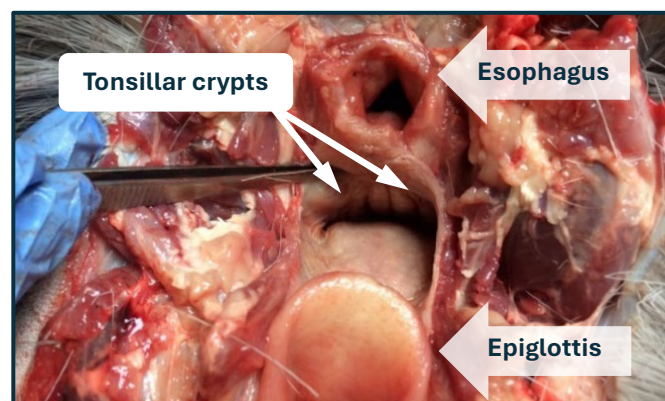
TIP: The lymph nodes will usually be pink, grey, or red in colour and are bean shaped. They can come in many sizes but are always contained within their own sac.

5. Carefully hold the very tip of the lymph node with forceps (tweezers).
6. Gently pull the lymph node and cut away the connecting tissue to extract it.
7. Collect the other lymph node and place both in a ziplock bag.



Tonsils

1. After extracting the lymph nodes, lift the trachea, pull towards the nose of the animal, and locate the epiglottis (firm cartilaginous flap).
2. Pull the epiglottis towards the nose of the animal and with a sharp knife, cut through the trachea into the back of the throat.
3. Locate the tonsillar crypts (openings or slits) on the upper area of the back of the throat. Each tonsil is located under the tonsillar crypts.
4. With forceps (tweezers), hold the crypt and use a knife to cut the tissue along the side of the throat, exposing the tonsil tissue.
5. Remove each tonsil and trim off any excess muscle tissue. Place both tonsils in the same ziplock bag as the lymph nodes.



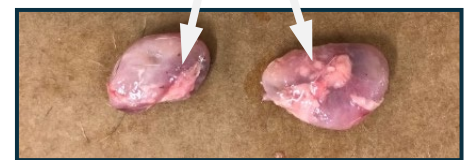
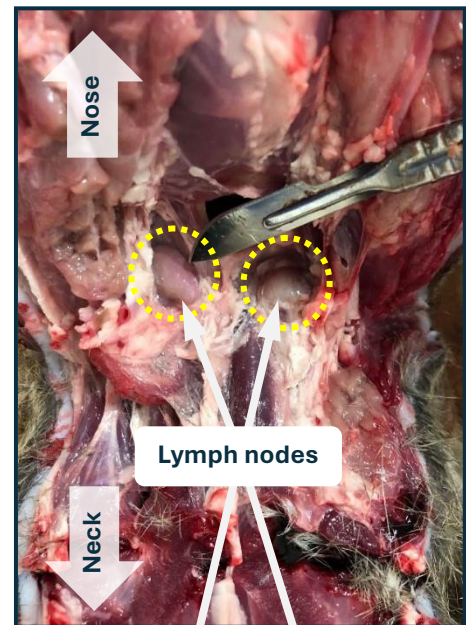
If you are sampling an **ELK**, **MOOSE**, or **CARIBOU**

Retropharyngeal Lymph Nodes (RPLNs)

1. Position head upside-down.
2. Using a sharp knife, cut underneath the jaw through the trachea and the muscles of the neck to the joint of the skull and the first vertebrae.
3. Lift the trachea, pull towards the nose of the animal, and cut away the muscles under the trachea until you reach fat pockets near the joint of the first vertebrae.
4. Cut through the fat pockets carefully to expose the left and right lymph nodes.

TIP: The lymph nodes will usually be pink, grey, or red in colour and are bean shaped. They can come in many sizes but are always contained within their own sac.

5. Carefully hold the very tip of the lymph node with forceps (tweezers).
6. Gently pull the lymph node and cut away the connecting tissue to extract it.
7. Collect the other lymph node and place both in a ziplock bag.



Obex

1. After extracting the lymph nodes, separate the skull from the first vertebrae. Cut the spinal cord as far away from the skull as possible.
2. Locate the brain stem at the base of the skull. Hold the membrane around the cord with forceps (tweezers) and carefully cut the cranial nerves with a sharp knife, scalpel, or scissors.
3. Carefully insert a spoon or scoop above the brainstem as far back in the skull as possible (until it contacts bone).
4. Tilt the spoon down to cut the tissue of the brainstem and gently remove the tissue from the skull.
5. Examine the brainstem to ensure the obex ("V") is intact. Place the entire obex in the same ziplock bag as the lymph nodes.



How to fill out your CWD Ear Card

CWD-24501 Ear Card

NAME: John Doe

PHONE: 250-123-4567

FWID #: 123 456 789

Mule Deer Elk Caribou
 White-tailed Deer Moose Black-tailed Deer
 Male Female

KILL DATE (dd/month/yyyy): 09/October/2024

KILL LOCATION: Cranbrook MU: 4 - 20

LAT/LONG or UTM: 49.53613, -115.758387

HOW KILLED: Hunted Motor Vehicle Collision
 Other (describe): _____

Was this animal normal? Yes No

If no, describe: _____

See reverse side for more information.

TEAR ALONG LINE AND RETAIN PORTION
OR TAKE PHOTO OF CARD TO LOOK UP RESULTS ONLINE

CWD-24501 Ear Card

NOTE:

The CWD Ear Card has been updated for 2024. You can still use older versions.

FRONT

Fill out the front side of the Ear Card and keep the perforated edge.

The unique code on the Ear Card can be used to look up your results online at

gov.bc.ca/CWDresults

KEEP!

Aim your phone camera at the QR code or visit our website at: www.gov.bc.ca/CWDresults to view your results when they become available.

B.C. Chronic Wasting Disease Program

For more information, contact B.C. Wildlife Health at 250.751.3219 or CWD@gov.bc.ca or visit the BC CWD Program website: www.gov.bc.ca/chronicwastingdisease

Instructions for head submission: Attach tag with a zip-tie (provided) to the ear or skull (if skinned). Place head with tag in a garbage bag (provided) and close with a knot before placing in freezer. **Do not attach Ear Card to outside of bag.**

If you are extracting tissues from the head, please fill out information below:

Date Sampled: 10/Oct/2024 Sampled by: Hunter Gov Staff Other: _____

Sample Condition: Good Hemorrhagic Fair European Mount: Yes No

Sample Type	Quantity	Reason If Not Collected
RPLNs (lymph node)	<input checked="" type="checkbox"/> 2 <input type="checkbox"/> 1.5 <input type="checkbox"/> 1 <input type="checkbox"/> 0.5 <input type="checkbox"/> 0	
Tonsils	<input checked="" type="checkbox"/> 2 <input type="checkbox"/> 1.5 <input type="checkbox"/> 1 <input type="checkbox"/> 0.5 <input type="checkbox"/> 0	
Obex (brain stem)	<input type="checkbox"/> Yes <input type="checkbox"/> No	
Ear tip	<input type="checkbox"/> Yes <input type="checkbox"/> No	Hunters not required to submit
Tooth (incisor)	<input type="checkbox"/> Yes <input type="checkbox"/> No	Hunters not required to submit
Tongue (in EtOH)	<input type="checkbox"/> Yes <input type="checkbox"/> No	Hunters not required to submit
Other:		Hunters not required to submit

Please leave blank for lab staff:

WLH ID: _____

Drop-off Location: _____

Data Entered:

BACK

If you are extracting samples from your own animal, please fill out the highlighted section on the back of your Ear Card.

As a hunter, you do not need to fill out these sections.